DISEASE RESPONSE STRATEGY

NEW WORLD SCREWWORM MYIASIS

FAD PReP

Foreign Animal Disease Preparedness & Response Plan

United States Department of Agriculture

United States Department of Agriculture • Animal and Plant Health Inspection Service • Veterinary Services

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USDA Animal and Plant Health Inspection Service (APHIS), Veterinary Services (VS)
Strategy & Policy ▪ National Preparedness and Incident Coordination

The continued spread of New World Screwworm (NWS) in Mexico and Central America has placed the United States (U.S). on high alert. APHIS has since taken several key steps to fortify the United States in the event of an NWS outbreak.

The *USDA Disease Response Strategy: New World Screwworm Myiasis (The Green Book)* provides an overview of current information on NWS and critical activities for response as guidance for development of Federal and State NWS disease response plans. The updated version of the *NWS Disease Response Strategy: The Green Book (June 2025)* reflects knowledge gained from policy discussions and response guidance development, previous NWS outbreaks in the United States and other countries, and lessons learned from Federal/State/Industry NWS preparedness activities. This version incorporates and supersedes previous versions of the *NWS Green Book*. Additionally, this version incorporates changes made in related Foreign Animal Disease Preparedness and Response Plan (FAD PReP) materials.

The following list highlights important revisions made to this version of the *NWS Disease Response Strategy:*

- Defines and clarifies the epidemiological principles for livestock and wildlife.
- Reflects updated case definitions and sample submission guidance.
- Briefly elaborates and clarifies federal, State, and tribal coordination from a One Health perspective.
- Refers operational and logistical details to response and guidance documents posted to FAD PReP.
- Provides and refers to newly posted list of EPA-approved pesticides reference for NWS to ensure validity.
- Clarifies active and passive surveillance activities.
- Corrects comments made and any errors identified in the prior version. Updates references throughout, as necessary.

While this *NWS Disease Response Strategy* provides strategic guidance before an outbreak, there are additional policy guidance materials in development and/or will be provided during an outbreak on specific response operation activities, particularly for the unified Incident Command. Please note certain topics have rapidly changing statuses. These types of issues may be referenced briefly in the *NWS Disease Response Strategy* but treated more fully in associated guidance documents. These additional policy guidance documents and information will be distributed and available at www.aphis.usda.gov/animal-emergencies/fadprep.

USDA APHIS acknowledges that preparing for and responding to an NWS outbreak is and will be a complex effort requiring collaboration and cooperation from all stakeholders. USDA APHIS fully anticipates updates as new capabilities and processes become available. As such, if you have comments or suggestions on this document, please send an email to FAD.PReP.Comments@usda.gov with the subject line: “Comments to Updated NWS Disease Response Strategy” for consideration and possible incorporation into future versions.

The FAD PReP mission is to raise awareness, define expectations, and improve capabilities for FAD preparedness and response. For more information, please go to www.aphis.usda.gov/animal-emergencies/fadprep or email *FAD.PReP.Comments@usda.gov*.

The Foreign Animal Disease Preparedness and Response Plan (FAD PReP)—*Disease Response Strategy: New World Screwworm Myiasis (2025)* provides strategic guidance for responding to an animal health emergency caused by New World screwworm (NWS) in the United States.

This *Disease Response Strategy: NWS Myiasis* was last updated in **June 2025**. Please send questions or comments to:

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While best efforts have been used in developing and preparing the *Disease Response Strategy: NWS Myiasis*, the U.S. Government, U.S. Department of Agriculture (USDA), Animal and Plant Health Inspection Service (APHIS), and other parties, such as employees and contractors contributing to this document, neither warrant nor assume any legal liability or responsibility for the accuracy, completeness, or usefulness of any information or procedure disclosed. The primary purpose of this document is to provide strategic guidance to those government officials responding to an NWS infestation. It is only posted for public access as a reference. Additional NWS disease response and guidance documents will be distributed and available at the FAD PReP website: www.aphis.usda.gov/animal-emergencies/fadprep with public materials posted by APHIS available here: www.aphis.usda.gov/livestock-poultry-disease/cattle/ticks/screwworm.

The *Disease Response Strategy: NWS Myiasis* may refer to links to various other Federal and State agencies and private organizations. These links are maintained solely for the user's information and convenience. If you link to such a site, please be aware that you are then subject to the policies of that site. In addition, please note that USDA does not control and cannot guarantee the relevance, timeliness, or accuracy of these outside materials. Further, the inclusion of links or pointers to particular items in hypertext is not intended to reflect their importance, nor is it intended to constitute approval or endorsement of any views expressed, or products or services offered, on these outside websites, or the organizations sponsoring the websites.

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INTRODUCTION

New World screwworms (NWS), *Cochliomyia hominivorax*, are fly larvae that infest living tissue of warm-blooded animals, causing a condition known as myiasis. Female NWS flies lay their eggs at the edges of wounds or on mucous membranes. The eggs hatch into larvae, which burrow into the tissue and continue to feed and grow. Infestations of NWS can be fatal if untreated. NWS is currently known to exist in parts of every country in South America except Chile, and in parts of the Caribbean. In 2023, Panama reported a drastic increase in the number of NWS cases. Since then, NWS has spread northward through the Central American countries and, in November 2024, reached the southern states of Mexico. NWS is not resident in the United States.

Other USDA APHIS Veterinary Services (VS) documents provide further detail on incident coordination and response to foreign animal diseases (FADs) and foreign pests, such as NWS. The *APHIS Foreign Animal Disease Framework: Roles and Coordination* (FAD PReP Manual 1–0) provides an introduction to APHIS FAD preparedness and response, an overview of the roles and responsibilities of different government agencies involved in an FAD response effort, as well as information on funding, incident management, and communication strategy. Additionally, an overview of FAD response strategies is available in the *APHIS Foreign Animal Disease Framework: Response Strategies* (FAD PReP Manual 2-0). These documents along with Disease Response Plans, National Animal Health Emergency Management System Guidelines, and other strategic documents are available publicly at www.aphis.usda.gov/animal-emergencies/fadprep.

NATURE OF THE DISEASE

NWS myiasis occurs when female flies are attracted to warm-blooded animals and lay eggs at the edges of wounds or on mucous membranes like nostrils, ears, eye orbits, mouth, or genitalia. Within 10 to 12 hours, larvae (commonly known as maggots) emerge from the eggs and immediately begin to feed. As they feed on host fluids and underlying tissues, the damage caused by their hook-like mouthparts enlarges and deepens the wound. The odor, serum, and blood emitted by the infested wound can attract other female flies that also lay their eggs; the result is additional myiasis and increasing damage. Severe infestations can lead to host death.

All living warm-blooded animals, including birds, can be infested by NWS, but it occurs most often in mammals (including humans). Unlike many other species of blow flies, female NWS flies will lay eggs only on living animals because NWS larvae do not feed on dead tissue or carrion.

Life Cycle

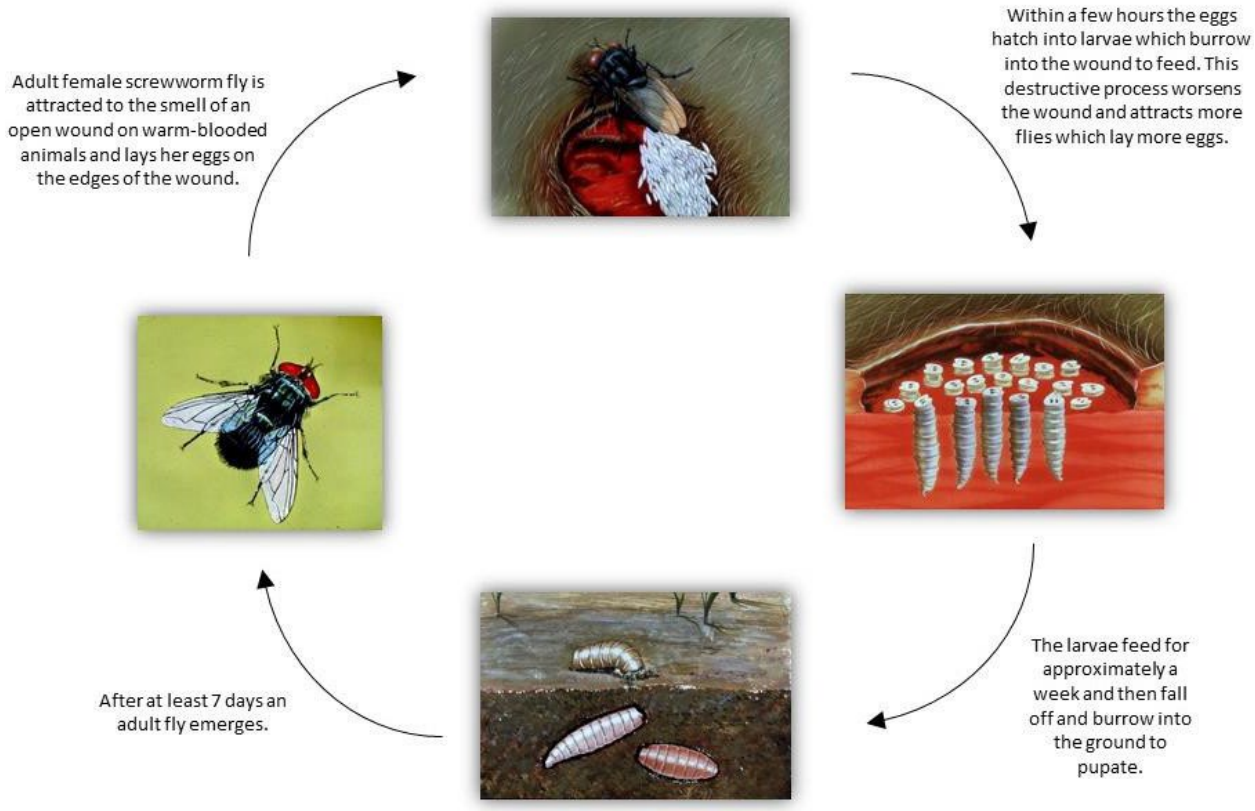
The duration of the NWS life cycle varies depending on the local climate. In cool climates, a complete life cycle may take 2 to 3 months, whereas in temperate climates (average temperature is 22° C/72° F), the NWS life cycle lasts approximately 24 days. In tropical conditions (29° C/84° F), the entire cycle may be as short as 18 days.

A female NWS fly mates with a male only once in her lifetime. Three to four days after mating she lays her eggs at the edge of an open wound or on mucous membranes of a living host. Every 3 or 4 days throughout her fertile lifespan, the female fly lays an egg mass of 100 to 350 eggs. The eggs hatch within 10 to 12 hours, and the larvae burrow into the wound and begin to feed. The larvae go through three stages of development, called instars, which involve two molts. The first molt (into the 2nd instar) occurs after about 24 hours, and the second molt (into the 3rd instar) occurs after an additional day.

Five to seven days after hatching, the mature larvae exit the wound and burrow a few centimeters into the ground where they pupate. If the larvae are unable to penetrate the soil or other substrate, they can move to a shaded area to pupate. Pupal development is temperature dependent. It can be as short as 7 days in warmer temperatures (28° C/82° F) or as long as 60 days in cooler temperatures (10–15° C/50–59° F), but pupae are killed if the soil is consistently below 8° C/46° F.

After emerging as adults, male flies live approximately 14 to 21 days and feed on flower nectar, while female flies live an average of 10 days (maximum 30 days) and feed on both flower nectar and on animal wounds as they deposit eggs. Males are ready to mate about 1 day after emergence, but females usually take 3 days to reach sexual maturity and become receptive to mating. Figure 1 provides an overview of the screwworm life cycle.

Figure 1. Life Cycle of New World Screwworm[1,2]



Morbidity and Mortality

Morbidity is variable for NWS myiasis; in areas with high NWS populations, the percentage of newborn animals with infested navel wounds can reach 100 percent. If left untreated, animals can die of trauma, toxicity, and/or secondary bacterial infections within 1 to 2 weeks. When NWS was still resident in South Texas in the 1950s, white-tailed deer fawns experienced mortality rates of 20 to 80 percent.

Clinical Signs and Differential Diagnosis

NWS myiasis is often associated with pre-existing wounds, though infestation can also occur on mucous membranes, such as nostrils, eye orbits, ears, mouth, and genitalia. Characteristics of infested wounds include drainage, suppuration (discharge of pus), discharge of blood and serum, enlargement, and distinctive

[1] Text adapted from: Fernández, P.J., & White, W.R. (2010). *Atlas of Transboundary Animal Diseases*. Paris: World Organization for Animal Health. 218–219.

[2] Source of photos: USDA APHIS STOP Screwworms: Selections from the Screwworm Eradication Collection, Special Collections, National Agricultural Library. http://specialcollections.nal.usda.gov/screwworm/index.

odor. Upon closer examination of the wounds, egg masses arranged in 'shingle-like' patterns/layers at the edges of the wound might be visible.

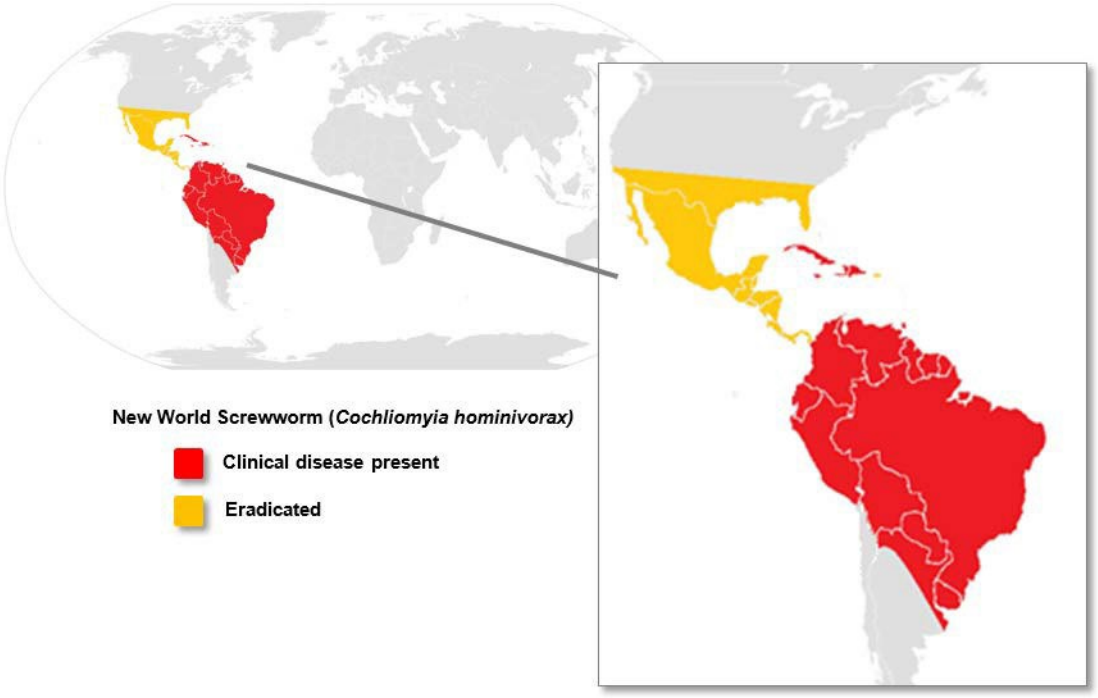
NWS larvae are visible by the third day; they can be 2.0 millimeters to 1.5 centimeters (0.08–0.60 of an inch) in length and are positioned with their posterior ends at the surface of the wound. In cases where the wound is deep, pocket-like and with a small opening, minor movement within the wound may be the only indicator of infestation. Secondary bacterial infections are also likely to occur, enlarging the wound area to 3 centimeters (1.2 inch) or more in width and up to 20 centimeters (7.9 inch) in depth. Co-infestation with maggot species that feed on dead tissue may also occur. Samples should be submitted to the USDA National Veterinary Services Laboratories (NVSL) for expert identification (see Laboratory Diagnosis).

Animals with NWS infestations often display discomfort, lethargy, depression, and may separate themselves from the herd. Anorexia and decreased milk yield may also be observed.

Occurrence

As illustrated in Figure 2, NWS has historically been found in tropical or semitropical regions of the Western Hemisphere, usually below 2,100 meters (7,000 feet) of elevation. From 2010 to 2016, the World Organisation for Animal Health (WOAH) received reports of NWS in either wild or domestic animals from Argentina, Aruba (immediately eradicated in 2011), Bolivia, Brazil, Colombia, Cuba, Dominican Republic, Guyana, Haiti, Jamaica, Panama (confined to zone near Colombian border), Paraguay, Peru, Suriname, Trinidad and Tobago, United States (eradicated in 2017), Uruguay, and Venezuela. NWS eradication programs have been successful in Curaçao, Puerto Rico, the U.S. and British Virgin Islands, United States, Mexico, and Central America, but a resurgence began in Central America in 2023 and spread northward to Mexico in 2024. Eradication efforts in these countries are on-going at the time of publication of this document. Imported cases of animal or human NWS myiasis are reported periodically in other countries where NWS has been eradicated.

Figure 2. Historical Distribution of New World Screwworm (pre-2023)



Environmental and Chemical Responses

NWS flies are able to travel long distances to find a suitable host. Although they generally remain within a 3-kilometer (1.9-mile) range when there is a high host density and suitable environment, flies are known to move 10 to 20 kilometers (6.2 to 12.4 miles) in warm, humid settings. Long distance movement of NWS is generally a result of the movement of an infested animal.

Although they prefer hot, humid environments, NWS are able to survive in a range of temperatures and climates. Table 1 provides information on the NWS response to certain environmental or chemical factors.

Table 1. Response of New World Screwworm to Physical and Chemical Factors[3]

Factor	Response
Soil Temperature	Pupae are killed in soil temperatures consistently below 8° C (46° F) or freezing.
Chemicals/Disinfectants	Organophosphate insecticides; carbamate and pyrethroid compounds.
Survival	Flies prefer hot, humid environments (air temperatures of 25–30°C with relative humidity of 30–70% is ideal) but can survive in suitable humid microclimates (e.g. irrigated areas) in otherwise dry conditions. In addition, availability of hosts with suitable wounds is fundamental.

Prevention

In areas where NWS is found, measures should be implemented to prevent wounds and avoid myiasis. For example, eliminate wounding procedures (e.g., castration), handle livestock with care, and inspect pens for sharp objects. Please see the Prevention of Wounds and NWS Myiasis section for more information.

Treatment

Organophosphate, carbamate, and pyrethroid compounds are effective against larvae and adult flies. Animals suspected of NWS infestation should have their wounds treated at 2- to 3-day intervals and remain in quarantine until the wounds are healed to ensure mature larvae are killed before leaving the wound and dropping on the ground to pupate. For more information, see the Treatment of Wounds and Myiasis section.

Laboratory Diagnosis

NWS myiasis is on the APHIS National List of Reportable Animal Diseases. Accredited veterinarians must collect, submit, and report suspect NWS myiasis. Diagnostic testing for NWS will be performed by the NVSL in Ames, Iowa. Although other agencies and entities may make a tentative identification, NVSL will assign the definitive parasite identification. Identification is by microscopic examination of the larvae (or possibly other life stages) and based on morphological characteristics. To obtain specimens for identification, gently remove larvae from several sites within the open wound using forceps. Because secondary myiasis may be present and is most often found near the surface of the wound, it is especially important to collect specimens from the deepest part of the wound. If possible, collect larvae of different sizes for submission.

[3] World Organisation for Animal Health (WOAH). (2013). Screwworm (Old World and New World). *Technical Disease Card.* www.woah.org.

Preserve the specimens in alcohol (ethyl or isopropyl), not formalin.[5] The concentration should be 70 percent or greater by volume. Specimens should be submitted to the NVSL with a Parasite Submission Form (VS Form 5-38) available on the APHIS website, located here: http://www.aphis.usda.gov/library/forms/pdf/VS_Form5_38.pdf.

For detailed information concerning the handling and shipping of diagnostic specimens, as well as overall guidance on FAD investigations, please see *APHIS VS Guidance Document 12001* and the *FAD Investigation Manual* (FAD PReP Manual 4-0), available at www.aphis.usda.gov/animal-emergencies/fadprep.

NWS Myiasis Response: Control and Eradication

The APHIS goals of an FAD response are to (1) detect, control, and contain the disease in animals as quickly as possible; (2) eradicate the disease using strategies that seek to protect public health and the environment, and stabilize animal agriculture, the food supply, and the economy; and (3) provide science- and risk-based approaches and systems to facilitate continuity of business for non-infested animals.

Achieving these three goals will allow individual livestock facilities, States, Tribes, regions, and industries to resume normal production as rapidly as possible. The objective is to allow the United States to regain NWS-free status without the response effort causing more disruption and damage than the outbreak itself.

Strategy for Responding to NWS Myiasis in the United States

Two basic epidemiological principles form the foundation of a response to control and eradicate an infestation of NWS in the United States.

1. *Prevent contact between NWS flies and non-infested animals.* This is accomplished through
    a. prevention, detection, and treatment of wounds in non-infested animals (domestic and wildlife);
    b. quarantine and treatment of infested livestock, movement controls, and inspections of livestock in regulatory control areas; and
    c. monitor and manage wildlife in affected areas.

2. *Stop the production cycle of NWS flies.* This can be accomplished through
    a. detection and appropriate treatment of infested animals; and
    b. use of the sterile insect technique (SIT) (see later section).

Case Definitions

Case Definitions for NWS can be found on the APHIS National List of Reportable Animal Diseases (NLRAD) website. The following case definitions were published in October 2023. In any NWS outbreak, case definitions may be revised after the first presumptive or confirmed positive case (index case). The case definitions will be reviewed throughout the outbreak and modified based on additional information or the changing needs of the eradication effort.

1. *Suspect case:* An animal with clinical signs consistent with blow fly or screwworm myiasis.

2. *Presumptive Positive Case:*

    a. *Imported case:* A suspect case that has travel history outside the United States to any screwworm-infested country within the previous 10 days.

    b. *Autochthonous case:* A suspect case that has no travel history outside the United States within the previous 10 days, **AND**

- located near a previous confirmed positive, **OR**
- identified as screwworm by any laboratory, **OR**
- identified as screwworm by a collector with screwworm experience.

3. *Confirmed Positive Case:*

    a. *Imported case:* A presumptive positive case in which the NVSL confirms the presence of screwworm by morphological identification of the egg mass, larva (first, second, or third instars), pupa, or adult fly.

    Autochthonous case: A presumptive positive case in which the NVSL confirms the presence of screwworm by morphological identification of the egg mass, larva (first, second, or third instars), pupa, or adult fly.

[5] USDA APHIS (2025) *Diagnostic Testing National Veterinary Services Laboratories - Ames, Iowa.* Parasite, external. http://www.aphis.usda.gov/animal_health/lab_info_services/downloads/AmesDiagnosticTestingCatalog.pdf.

Reporting

Under the APHIS NLRAD, New World and Old World screwworm myiasis are reportable foreign animal diseases (FAD) in the United States and a WOAH-notifiable disease. Suspect cases must be reported to a State Animal Health Official (SAHO) or Area Veterinarian in Charge (AVIC) who will decide if the report is credible and assign a Foreign Animal Disease Diagnostician (FADD) to further investigate the possibility of NWS detection or infestation. For more information on FAD investigation procedures please refer to VS Guidance Document 12001.5: Policy for the Investigation of Potential Foreign Animal Disease/Emerging Disease Incidents and the FAD Investigation Manual (FAD PReP Manual 4-0), available at https://www.aphis.usda.gov/animal-emergencies/fadprep.

Critical Activities

Epidemiology Investigation and Tracing

For any detection of NWS myiasis in the United States, a thorough epidemiological investigation is required to determine if the infestation was acquired domestically or in a foreign location. If it is concluded that the myiasis was not acquired in the United States, the investigation must determine if a domestic population of NWS may be in the process of developing due to the myiasis introduced from a foreign source. If a domestic population of NWS is known to be present or suspected to be developing, epidemiological investigation and movement tracing are critical in controlling and eradicating that population.

Epidemiological investigation and tracing are typically the responsibility of multiple staff components within the Incident Command System (ICS), including the Disease Surveillance Branch (Operations Section), Epidemiologists (Planning Section), and the National Situation Unit (Planning Section).

Tracing

Trace Back: Identifying the origin and movements over the last 10 days for all animals and people detected with myiasis, in order to determine the original source of the myiasis, or potential locations of additional infestation.

Trace Forward: The tracing of all animals and people that have left an infested location in the last 10 days and could have possibly transmitted infestation to a new location. The premises that received the animals should be investigated and kept under surveillance.

Information Management

Information management and reporting during an NWS outbreak ensures that responders, stakeholders, and decision-makers have access to accurate and timely critical emergency response information. Ideally, local, State, Tribal, and Federal information management systems are compatible for information and data

sharing. The Emergency Management Response System (EMRS) is the official USDA APHIS system of record during an NWS outbreak. EMRS contains data on infested animals, fly releases, fly trapping, movements and traces, any zones established, and other information.

In an NWS outbreak, the response goal is to have EMRS information downloads or data entry processes performed at 12-hour or shorter intervals. Data should be entered as quickly as possible in both an accurate and consistent manner across widespread field operations. Field personnel should be provided with access to mobile technology devices necessary for collecting, monitoring, and sharing information. Information within EMRS will provide the data necessary for daily, weekly, and ad-hoc situation reports. It is imperative that outbreak information management, data quality, and data integrity are a priority. Additional resources on information management systems and tools can be found in *Information Management and Reporting* (FAD PReP Manual 3-0).

Communication and Coordination

APHIS Legislative and Public Affairs (LPA) will serve as the primary Federal government liaison with the news media in the event of an NWS outbreak. Under the ICS, a Joint Information Center (JIC) is established. During an NWS outbreak, APHIS LPA and the USDA Office of Communications operate from the JIC.

Effective communication should be carried out and maintained by

- establishing a network of stakeholders and systems for communication prior to an incident or outbreak;
- briefing the media, public, industry, Congress, trading partners, and others on the NWS outbreak status and the actions being taken to control and eradicate the infestation;
- assuring consumers that USDA is working on animal health issues and the Department of Health and Human Services (HHS) is working on human health issues, in an informed and timely manner, along with other federal counterparts:
    - the Department of Health and Human Services (HHS) Centers for Disease Control and Prevention (CDC) and Food and Drug Administration (FDA), the Department of Homeland Security (DHS), the Department of the Interior (DOI), the Environmental Protection Agency (EPA), as well as
    - state, local, tribal, and territorial (SLTT) partners from animal health (domestic and wildlife), public health, and environmental agencies.

NWS can infest domestic animals, wildlife, and in rare cases, humans. Responding to a potential introduction of NWS in the United States will require a collaborative, multisectoral, and transdisciplinary approach with collaboration across the animal, human, and environmental health sectors.

In addition, all communications should highlight the importance of sound

preventive measures and steps that producers and owners can take to protect against NWS infestation in their own animals.

Public Education

Educating the public, particularly animal owners and hunters, to recognize and report possible cases of myiasis will be critical. A large publicity campaign may be warranted to broadly disseminate information and seek public cooperation during an NWS response. Due to the wide host range for NWS, awareness campaigns should partner with public health and wildlife agencies.

Providing timely information to local livestock owners and producers, veterinarians, physicians, hunters, pet owners, and the general community can help generate reports of possible cases of NWS myiasis. Education on wound prevention and treatment can help limit the number of cases that occur. Cooperative Extension System agents have contact with animal owners and can serve a valuable role in an educational campaign. Although educational material may need to be tailored to the location and population affected by an outbreak, a basic example of information for the public is available at https://www.aphis.usda.gov/publications/animal_health/bro-new-world-screwworm.pdf.

Additional APHIS FAD PReP resources for the public include NWS Ready Reference Guides (1) Maps and Timelines, (2) Historical Economic Impact, and (3) Sterile Insect Response. Also available is the NWS Story Map, which includes interactive maps on the 2016–2017 Florida NWS Response. Links to these materials are available in the References and Resources section.

Quarantine and Movement Control

Quarantine refers to imposing restrictions on entering or leaving a premises, area, or region where disease exists or is suspected. Quarantine stops the movement of infested animals.

Movement control refers to activities regulating the movement of animals within an area subject to certain criteria. Movement control is accomplished through a permit system that allows entities to make necessary movements without creating an unacceptable risk of spreading the infestation.

Quarantines and movement controls are effective measures to prevent the spread of infestation. While controlling the spread of female NWS flies is nearly impossible, dispersal has been observed to be largely due to movement of animals with myiasis and not the result of flies seeking out new hosts.[6] These animal movements can result in long-distance spread and establishment of infestations in entirely new geographic areas. Therefore, quarantines and movement controls will be a primary strategy of NWS response efforts.

USDA may impose a Federal area quarantine and restrict interstate commerce from the infested States, asking the States (or adjoining countries) to provide resources to maintain and enforce the quarantine. State quarantines may be placed on individual infested animals or premises with infested animals. Of most importance is the control of livestock movement within and out of an infested area, using a system that requires inspection for wounds and myiasis. This may include permit requests for permitted movement.[7] For example, ICS personnel could issue a permit for movement out of a quarantine area after an inspection for wounds and myiasis, followed by treatment of the animal with an approved insecticide. Other approaches may also be reasonable, based on the epidemiological characteristics of the incident.

All decisions in regard to quarantine and movement control will be based on science-based assessments of the current extent of NWS infestation, risk of spread, and the interaction of other factors, such as seasonal climate and weather conditions.

Control Area (or Other Zone) Designations

Appropriate Control Area (or other zone) designations may be required for implementation of quarantine and movement control measures. The Incident Commander will work with the Disease Surveillance Branch (Operations Section) and Situation Unit (Planning Section) to establish a Control Area or other zones, once it is determined that a domestic population of NWS is present or suspected to be developing. Once the Control Area is established, quarantine and movement controls, including a permit system (as appropriate), may be implemented.

NWS Eradication Using the Sterile Insect Technique (SIT)

SIT employs exposure of lab/factory-reared 5- to 6-day-old NWS pupae to gamma radiation to create sterile NWS files. Field release of these mass-produced sterile flies results in sterile males overwhelmingly mating with wild female flies that then lay unfertilized eggs. Because female NWS flies normally mate only once, the wild population of NWS is progressively reduced and, ultimately, eradicated. SIT does not involve the release of insects modified through genetic engineering processes. The release of sterile flies is safe, environmentally friendly, and offers a sustainable, non-toxic alternative to chemical pesticides. It poses no risk to wildlife, livestock, or people in infested areas.

USDA scientists performed a successful SIT field experiment in 1951 on Sanibel Island, Florida, but its proximity to infested mainland Florida didn't allow for long-term eradication. In 1954, on the Caribbean Island of Curaçao (Netherlands Antilles), SIT was used effectively to eradicate NWS in approximately 6 months. The next set of SIT applications in the United States started in 1957 in Florida. After the success of this campaign, SIT was used to eradicate NWS successively from the United States, Mexico, and Central America.

Before the most recent autochthonous cases of NWS myiasis were detected in Florida in 2016, there had been at least 27 post-eradication findings of NWS larvae in animals or people in the United States.[8,9,10,11] These cases were typically linked to recent travel or residence of the host in an NWS-endemic country. In two separate incidents in 1987, both involving dogs with NWS myiasis detected after return from foreign travel, it was determined that use of the SIT was warranted.

Sterile flies were released for 6 weeks around locations in Florida, Louisiana, and New Mexico, and no additional local NWS larvae were ever found.

Use of the Sterile Insect Technique

Use of the SIT should be considered if it is determined that a population of NWS is present, or may be in the process of developing, within the United States. This determination is made based on multiple factors, or combinations of factors, relative to the index case of myiasis detected in an animal or human host. These factors include: stage of larval development, the affected host's history of international travel, time elapsed since United States entry, locations visited since returning, treatment provided, and environmental and seasonal considerations.

More specifically, there are two situations in which SIT should be considered:

1. finding NWS larvae in an animal or person that did not travel outside of the United States in the previous 10 days;
2. finding NWS larvae in an animal or person that did travel outside of the United States in the previous 10 days, with evidence that larvae may have left the host in the United States (e.g., myiasis detected 5 or more days after United States arrival, or collection of NWS pupae) and could complete pupation (e.g., access to soil, favorable local temperatures, suitable seasonal climate).

In the second situation, intensive surveillance for NWS myiasis or flies may initially be a reasonable alternative to using the SIT. If surveillance finds evidence that confirms a population of NWS is present in the United States, SIT would then be called for to pursue eradication.
It is important to note that the SIT would be just one of several control strategies employed in the event of an infestation; movement controls, tracing, surveillance, and treatment and prevention would be components of any NWS myiasis response.

Sterile Fly Production

APHIS International Services (IS) maintains the only NWS pupal sterilization facility in North America. The facility, located in Pacora, Panama, is jointly managed and funded by USDA and Panama's Ministry of Agriculture Development through the Screwworm Barrier Maintenance Program (also known as COPEG). COPEG produces, sterilizes, and releases NWS sterile flies in infested regions of Central America and Mexico. The COPEG facility can produce over 100 million pupae per week during an outbreak event. USDA is working on increasing the availability of

sterile flies to meet the current demands of the NWS outbreak by opening additional sterilization facilities.

For additional details, refer to this factsheet, Eradicating New World Screwworm with Sterile Insect Technique which can be found on the public APHIS Screwworm website: www.aphis.usda.gov/livestock-poultry-disease/cattle/ticks/screwworm.

Timing of Sterile Fly Release

In a worst-case scenario, the initial detection of NWS myiasis would not occur until 3 weeks or more after the initial introduction of NWS larvae into a United States location favorable for pupation and ongoing maintenance of the NWS life cycle. In this scenario, a viable population of NWS flies would already exist in the United States at the time of myiasis detection in an animal or person. Releases of sterile NWS flies would be required for at least 9 to 12 weeks in order to control and eventually eradicate that population. The sterile fly releases would need to begin as soon as possible. During the 2016–2017 NWS outbreak in Florida, samples were not sent to the NVSL for identification until 84 days (4 life cycles) after presumptive positive identification by local veterinarians.

A more likely scenario in which the initial introduction of myiasis is detected in an animal or person that recently entered the United States from an NWS-infested country, and it is determined that some of the larvae may have already exited the wound and begun to pupate. In such a scenario, even under ideal climatic conditions, it would be at least 10 days after the host entered the United States before the first female NWS flies could be mature and ready to mate. If release of sterile male NWS flies began by that time, establishment of a population of NWS flies probably could be prevented.

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[6] Animal Health Australia. (2007). *Disease Strategy: Screw-Worm Fly* (Version 3.0). Australian Veterinary Emergency Plan, Edition 3, Primary Industries Ministerial Council, Canberra, ACT.

[7] For more information on permitting, please see the document *Permitted Movement* (FAD PReP Manual 6-0). [8] Alexander, J.L. (2006). Zoonosis Update: Screwworms. *Journal of the American Veterinary Medical Association*, 228(3), 357-367.

[9] Spickler, A.R., & Roth, J.A. (Eds.). 2006. Screwworm-New World (*Cochliomyia hominivorax)* in the United States, 2000. In *Emerging and Exotic Diseases of Animals* (3rd ed.: 105-107). Ames, IA: Institute for International Cooperation in Animal Biologics, Iowa State University College of Veterinary Medicine.

[10] USDA APHIS. (2008). 2007 United States Animal Health Report. *Agriculture Information Bulletin No. 803*. http://www.aphis.usda.gov/animal_health/animal_health_report/downloads/AHR_08/ahr2007.pdf.

[11] Corn, J.L. (Ed.). (2011). *Proceedings - 114th Annual Meeting of the United States Animal Health Association*. Saint Joseph, MO. http://www.usaha.org/upload/Proceedings/USAHAProceedings-2010-114th.pdf.

Example of Release Logistics

Each NWS incursion or outbreak situation will be evaluated by APHIS subject matter experts and a specific plan will be formulated for dispersal considering location, habitat, method of dispersal, and other logistics. These plans will take into consideration factors described in this section.

Pupae would be shipped by express air delivery in large polystyrene foam coolers or insulated chests. Due to variability over time, pupal size will be continually monitored to better estimate the quantity of sterile pupae that are shipped.

The simplest and most rapidly implemented dispersal of sterile NWS flies will be through ground release. Ground release containers were used in Chiapas, Mexico, in outbreaks in 2001 and 2003, to augment aerial dispersal of sterile flies in specific key areas. Ground release containers were the sole means of dispersing sterile flies to eradicate outbreaks in Aruba in 2004 and 2011, and in Florida during the 2016–2017 outbreak.

Ground release containers may be cardboard or plastic boxes, or even larger structures, depending on the needs of the situation. Release containers (Figure 3) can be placed in the field as early as the day following arrival from Panama, or the emergence of the flies can be delayed for a few days by chilling the pupae to 50º F (10 º C)

Figure 3. Release Container Used in the Florida Keys, 2016–2017

Photo: USDA APHIS Veterinary Services

Sterile NWS flies would be released throughout an area within 10 kilometers (6.2 miles) of each location where a case of myiasis has been detected. Similarly, in cases where an infested animal or person traveled to multiple locations during the period of larval maturation, release of flies may be warranted within 10 kilometers of each location where pupation could have occurred.

Alternatively, in such exposed areas where myiasis has not been detected but fly development is considered possible, intensive surveillance may be applied initially, and sterile fly release may begin only if NWS myiasis is subsequently detected or wild NWS flies are trapped in the area.

Dispersal of flies by air, from specially equipped fixed-wing aircraft, should be considered if factors, such as the large size of an infested area or limited accessibility to large portions of an infested area, make ground release impractical or ineffective.

Throughout the dispersal area, the weekly release rate of sterile NWS flies should be 875 per square kilometer, or 2,266 per square statute mile, or 3,000 per square nautical mile.

With ground release of flies, the pattern used in air release could be duplicated or some other pattern that achieves a uniform dispersal over a 7-day period could be employed. Depending on their size, release containers may be filled with from 1,000 to 240,000 pupae each; this will produce a minimal release of 850 to 204,000 sterile flies per container placement. A practical and versatile release container might contain 2,000 pupae to produce at least 1,700 sterile flies for release. Such a container would provide coverage for an area of about 4 square kilometers, or about 1.5 square statute miles, for half a week (i.e., each square kilometer needs 875 flies per week, each square statue mile needs 2,266 flies per week); the proper placement of 100 of these containers, twice a week, would release approximately enough flies to cover one complete 400 square kilometer infestation area.

Another advantage of ground release is that it may be possible to more closely target habitats that are favorable for wild NWS flies rather than simply using a uniform dispersal pattern. Analysis of satellite imagery of the intended release area can potentially locate favorable habitat in which to concentrate ground releases. If favorable habitat areas are accessible for the placement of fly containers, more than 875 flies per square kilometer would be effectively released each week, and fewer flies would be released in areas with less favorable habitats. Experts from APHIS IS, APHIS VS, and the Agricultural Research Service of USDA should be consulted for such analysis.

[14] Smith, S.C. (2009). A Guide for the Eradication of Screwworms. Comision Mexico - Americana para la Eradicacion del Gusano Barrenador. http://www.flsart.org/screwworm/Screwworm%20Eradication%20Guide%20Print%20Version.pdf

Weekly Release Rate Summary

- Per square kilometer: 875 sterile NWS flies
- Per square statute mile: 2,266 sterile NWS files
- Per square nautical mile: 3,000 sterile NWS flies

Example

For an *infestation area* of 400 square kilometers, the following would result in a sufficient release of sterile NWS flies:

- 100 containers
- Placed twice per week
- 2,000 pupae per container (with an emergence rate of 85 percent or more, this results in at least 1,700 sterile flies)

The area of release may be enlarged or reduced based on local conditions, including distributions of livestock, wildlife, and human populations. To facilitate the logistics of release, particularly by air, it may be useful to square the 10 kilometer radius around an infested location into a 20 kilometers by 20 kilometers (12.4 miles by 12.4 miles) square area, comprising a total of 400 square kilometers (approximately 154 square miles) and centered on the location of myiasis detection or possible pupation.

Release of sterile flies should continue for at least three life cycles, or about 9 weeks in warm conditions, past the last detection of NWS myiasis within any given circle of 10 kilometer radius. Continuing release for four life cycles (about 3 months) would provide even more certainty of eradication. Factors such as local weather, seasonal climate, and ecological conditions should be considered in determining when to stop releasing sterile flies.

SURVEILLANCE

The purpose of surveillance for NWS is to define the extent of any current infestation, detect new infestations, support establishment of NWS-free areas, and demonstrate effectiveness of disease control efforts. Surveillance is also used to support continuity of business and provide evidence for freedom from NWS myiasis following an outbreak. These objectives will be achieved by comprehensive surveillance plans incorporating active and passive surveillance of domestic animals, surveillance in wildlife, and surveillance for NWS flies.

Visual and olfactory inspection of animals will be the primary surveillance tool for animal infestations. Animals on premises in response areas that are at risk for NWS exposure will be regularly surveyed for evidence of NWS myiasis. The frequency of surveillance will be informed by the lifecycle of NWS flies, epidemiology and pathogenesis of disease, and response objectives. Animals on premises in the control area must be visually inspected for the lack of wounds and myiasis as a component of permitted movement within, and potentially out of, the control area. All maggot specimens detected must be submitted to the NVSL for definitive identification.

Passive surveillance will be critical to support response and eradication. Educating local livestock owners and producers, veterinarians, physicians, hunters, wildlife rehabilitators, biologists, pet owners, and the general community will facilitate passive surveillance for NWS myiasis.

Due to the wide host range for NWS, awareness campaigns will also partner with public health and wildlife agencies. Although the confirmation of NWS must be made by the NVSL, possible cases of myiasis in people will be reported through public health agencies; partnering with Federal and State wildlife agencies will facilitate surveillance for NWS myiasis in wildlife species.

Surveillance for NWS flies should be used to determine the geographic spread of flies, document the absence of fertile NWS flies (through their absence in traps) in the zone(s) surrounding a control area, and to provide evidence of freedom from NWS after sterile fly release is discontinued. Trapping may also be used within the response areas to evaluate the release, dispersal, and efficacy of sterile NWS flies.

Methods available for trapping NWS flies include luring flies and collecting them with nets, wind-oriented traps with chemical attractants or baits, including sticky fly strips. Traps may be set up at varying distances from points of known infestation and should account for typical flight distance of NWS flies and weather patterns. Trapping and surveillance activities will be conducted in favorable NWS habitats, such as the edges of wooded areas, and in such a manner that the trapping/surveillance site is located up-wind, so the odor of the attractant carries into the favorable habitat. For more information, please see the document *Fly Surveillance & Site Selection Methods* (www.aphis.usda.gov/animal-emergencies/fadprep, navigate to the NWS page). More detailed guidance on designing an outbreak-specific surveillance plan and conducting surveillance activities in the field will be available to Disease Surveillance Branch and other Incident Command personnel during an incident.

Surveillance data from all activities (e.g. active domestic animal surveillance, passive surveillance, wildlife surveillance, fly surveillance, etc.) should be integrated for analysis, visualization and timely communication to response agencies and stakeholders. See section *Information Management* for more details.

Treatment of Wounds and Myiasis

All wounds detected in animals within a control area should be treated prophylactically with insecticide, every 2 to 3 days until healed, to prevent female NWS flies from laying eggs and to kill any eggs or larvae already present. All detected myiases must be treated. The protocol for treatment is as follows:

1. Clean the wound and the surrounding tissue with warm water and a mild antiseptic.
2. If myiasis is found, physically remove as many larvae as possible. Retain 10 or more larvae for identification (see Laboratory Diagnosis section).

3. Apply an effective topical treatment to kill any remaining larvae.
4. Treat animal with longer-lasting systemic insecticide for prophylaxis.

The List of Pesticides to Potentially Use Against New World Screwworm provides current Environmental Protection Agency (EPA)-registered products with known efficacy against NWS. All products are registered only for labeled use. Please check the EPA website (https://www.epa.gov/pesticides) to determine if products are still registered for use.

Prevention of Wounds and NWS Myiasis

Within the Control Area, measures should be implemented to prevent animal wounds and avoid NWS myiasis. For example, to the extent possible, eliminate or delay performing wounding procedures such as dehorning, branding, shearing, ear notching, tail docking, and castration. Additionally, livestock should be handled with care, and pens should be inspected for sharp objects. Measures should be taken to protect livestock from other wound-causing parasites, such as ticks. Untreated umbilical cords of newborn animals and foot lesions are commonly infested sites. Immediately treat all wounds with approved insecticides; it may also be prudent to follow up with precautionary spraying of animals with insecticide before transport.

Wildlife Management

It should be assumed that some wildlife within an infested area will be subject to NWS myiasis. It is therefore important to minimize the potential dispersal of wildlife located within an NWS-infested area. One measure that should be considered is a temporary prohibition on hunting in the control area. APHIS VS will collaborate closely with Federal, State, and local wildlife health officials to ensure that the most appropriate decisions are made in managing wildlife.

Euthanasia

During NWS surveillance and control, individual animals may require euthanasia for welfare reasons.

Disposal

Disposal frequently is not a great concern, as depopulation activities will typically be minimal. If small numbers of dead animals must be disposed of, responders must ensure that no viable NWS larvae are on or in the carcass, especially if burial is used. Incineration is a preferred disposal method for NWS-infested carcasses. Because NWS feed only on living flesh, the focus of pre-disposal examinations should be on very recently deceased animals. For further information on disposal, see the *National Animal Health Emergency Management System (NAHEMS) Guidelines*: *Disposal* available at www.aphis.usda.gov/animal-emergencies/fadprep.

Cleaning and Disinfection

Because NWS larvae are obligate parasites, requiring living flesh to remain viable, it is extremely unlikely that larvae would be transported on animal products or byproducts. However, there is a small risk that carcasses of very recently deceased animals might still be infested, and they should therefore be thoroughly checked before leaving the control area. Be aware that larvae typically and rapidly vacate a wound and pupate when the host animal dies.

NWS larvae normally pupate in 2 to 3 centimeters (about 1 inch) of soil, but pupation can occur anywhere there is a slight buildup of organic material, such as in feces. Maggots can also pupate in other protected sites, like cracks and crevices, or below bedding materials. Therefore, areas where infested or susceptible animals have been held, including vehicles, must be immediately and thoroughly cleaned and treated with insecticide as part of an effective control effort. Furthermore, animal material that may contain viable pupae, such as manure, should be disposed of in a manner that will conclusively kill that stage; incineration is preferred. To avoid these situations, inspections and cleanings of transport vehicles should be conducted in areas with concrete or otherwise hard and sealed floors. Trucks, trailers, or other conveyances used to transport infested animals should also be treated with insecticide before further use.

Health and Safety and Personal Protective Equipment

Protecting the health and safety of personnel assigned to emergency response activities is everyone's responsibility. During an outbreak response, responders may be exposed to a number of different hazards. Taking precautions to prevent adverse human health events related to emergency response efforts is important.

NWS larvae can infest humans, so responders should take precautions when potentially exposed to NWS flies. If responders have open wounds, those injuries should be thoroughly cleansed and covered. For most responders, the appropriate level of personal protective equipment (PPE) during screwworm response efforts will be Level D. Level D is the lowest level of protection and requires only a basic work uniform to protect the body against contamination and no respiratory protection. Disposable gloves and possibly cut-resistant gloves (if warranted by specific activities), are needed for hand protection. Although this is the suggested minimal level of PPE for screwworm, the final decision is always based on an assessment of the specific circumstances and activities in the field. Some response activities may require the use of protection higher than Level D.

For further information on health, safety, and PPE, associated with general response hazards, see the *NAHEMS Guidelines: Health and Safety and Personal Protective Equipment* available at www.aphis.usda.gov/animal-emergencies/fadprep. For additional information on NWS infestation prevention in humans, see CDC's website at www.cdc.gov/myiasis/about-new-world-screwworm-myiasis/index.html#cdc_generic_section_5-prevention.

Indemnity and Compensation

The response to an NWS infestation should not require significant, if any, destruction of animals or other assets. However, USDA is authorized by the Animal Health Protection Act (7 U.S.C. 8301 et seq.) to pay claims to owners for any assets taken or destroyed in the course of a response effort. Title 9 of the Code of Federal Regulations, Part 53, outlines the expenses that the Department may pay for purchasing, destroying, and disposing of animals and materials in these situations. Fair market value appraisals will be made for animals and materials destroyed to prevent the spread of NWS. Please refer to the Producer Indemnity and Compensation website for further information.

Criteria for Proof of Freedom from NWS Infestation

WOAH does not provide specific criteria for proof of freedom from NWS myiasis. A declaration of NWS freedom after an infestation in the United States would be based on surveillance for NWS that includes trapping of flies and visual inspection of animals for myiasis. Depending on specific weather and seasonal conditions in the area(s) of infestation, surveillance should be carried out for 3 months past the last NWS detection, generally 3–4 life cycles (~24 days/life cycle) during an outbreak. Historically, when eradicating NWS from a previously endemic area, USDA APHIS declares an area to be "Technically Free" of NWS after 6 months without a detection and "Officially Free" after 12 months without a detection.

REFERENCES AND RESOURCES

Alexander, J.L. (2006). Zoonosis Update: Screwworms. *Journal of the American Veterinary Medical Association*, 228(3), 357-367.

Animal Health Australia. (2007). *Disease Strategy: Screw-Worm Fly* (Version 3.0). Australian Veterinary Emergency Plan, Edition 3, Primary Industries Ministerial Council, Canberra, ACT.

Baumhover, A.H. (2002). A personal account of developing the sterile insect technique to eradicate the screwworm from Curacao, Florida, and the Southeastern United States. *Florida Entomologist*, 85(4): 666-673.

Center for Food Security and Public Health, Iowa State University. (2012). "Screwworm Myiasis Technical Fact Sheet. http://www.cfsph.iastate.edu/Factsheets/pdfs/screwworm_myiasis.pdf.

Concha, C., Palavesam, A., Guerrero, F.D., Sagel, A., Li, F., Osborne, J.A., … Scott, M.J. (2016). A transgenic male-only strain of New World screwworm for an improved control program using the sterile insect technique. *BMC Biology,* 14(72): DOI 10.1186/s12915-016-0296-8.

Corn, J.L. (Ed.). (2011). *Proceedings - 114th Annual Meeting of the United States Animal Health Association*. Saint Joseph, MO. http://www.usaha.org/upload/Proceedings/USAHAProceedings-2010-114th.pdf.

Fernández, P.J., & White, W.R. (2010). *Atlas of Transboundary Animal Diseases*. Paris: World Organization for Animal Health. 218-219.

Gutierrez, A.P., & Ponti, L. (2014). The new world screwworm: prospective distribution and role of weather in eradication. *Agricultural and Forest Entomology,* 16(2), 158-173.

Mastrangelo, T., Chaudhury, M.F., Skoda, S.R., Welch, J.B., Sagel, A., & Walder, J.M.M. (2012). Feasibility of using a Caribbean Screwworm for SIT Campaigns in Brazil. *Journal of Medical Entomology,* 49(6), 1495-1501.

Mastrangelo, T., & Welch, J.B. (2012). An Overview of the Components of AW-IPM Campaigns against the New World Screwworm. *Insects,* 3, 930-955.

National Veterinary Services Laboratories—Ames. (2025). Diagnostic Testing. http://www.aphis.usda.gov/animal_health/lab_info_services/downloads/AmesDiagnosticTestingCatalog.pdf.

Novy, J.E. (1991). Screwworm control and eradication in the southern United States of America. *World Animal Review,* FAO Special Issue. http://www.fao.org/docrep/U4220T/u4220T0a.htm.

Smith, S.C. (2009). A Guide for the Eradication of Screwworms. Comision Mexico - Americana para la Eradicacion del Gusano Barrenador. http://www.flsart.org/screwworm/Screwworm%20Eradication%20Guide%20Print%20Version.pdf

Scott, M.M., Concha, C., Welch, J.B., Phillips, P.L., & Skoda, S.R. (2017). Review of advances in the screwworm eradication program over the past 25 years. *Entomologia Experimentalis et Applicata,* Special Issue, 1-11.

Spickler, A.R., & Roth, J.A. (Eds.). (2006). Screwworm-New World (*Cochliomyia hominivorax)* in the United States, 2000. In *Emerging and Exotic Diseases of Animals* (3rd ed.: 105-107). Ames, IA: Institute for International Cooperation in Animal Biologics, Iowa State University College of Veterinary Medicine.

USDA APHIS. (2017). Cooperative Screwworm Eradication Program Environmental Assessment. https://www.aphis.usda.gov/sites/default/files/screwworm-environmental-assessment.pdf.

USDA APHIS. (2008). 2007 United States Animal Health Report. *Agriculture Information Bulletin No. 803*. http://www.aphis.usda.gov/animal_health/animal_health_report/downloads/AHR_08/ahr2007.pdf.

USDA APHIS. STOP Screwworms: Selections from the Screwworm Eradication Collection. Special Collections, National Agricultural Library. http://specialcollections.nal.usda.gov/screwworm/index.

USDA APHIS. (2025). New World Screwworm. https://www.aphis.usda.gov/animal-emergencies/nws.

USDA APHIS. (2017). New World Screwworm Story Map. https://www.aphis.usda.gov/aphis/maps/animal-health/screwworm-mapping.

USDA APHIS. (2018). New World Screwworm: What You Need to Know. https://www.aphis.usda.gov/publications/animal_health/bro-new-world-screwworm.pdf.

Vargas-Terán, M., Hursey, B.S., & Cunningham, E.P. (1994). Eradication of the screwworm from Libya using the sterile insect technique. *Parasitology Today,* 10(3), 119-122.

World Organization for Animal Health. (2013). Screwworm (Old World and New World). *Technical Disease Card.* www.woah.org.

Wyss, J.H. (2000). Screwworm Eradication in the Americas. *Annals of the New York Academy of Sciences,* 916, 186-193.

ABBREVIATIONS

APHIS	Animal and Plant Health Inspection Service
CEAH	Center for Epidemiology and Animal Health
EMRS	Emergency Management Response System
EPA	Environmental Protection Agency
FAD	foreign animal disease
FAD PReP	Foreign Animal Disease Preparedness and Response Plan
ICS	Incident Command System
IS	International Services
JIC	Joint Information Center
LPA	Legislative and Public Affairs
NAHEMS	National Animal Health Emergency Management System
NVSL	National Veterinary Services Laboratories
NWS	New World screwworm
WOAH	World Organisation for Animal Health
PPE	personal protective equipment
SIT	sterile insect technique
U.S.C.	Code of Laws of the United States of America
USDA	United States Department of Agriculture
VS	Veterinary Services